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**StressXpress®**

## **Glutathione S-Transferase Activity Kit**

Catalog# SKT-203 (96-Well Kit)

Quantitative fluorometric measurement of the activity of GST



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## GENERAL INFORMATION

### Materials Supplied

Catalog Number	Reagent	Quantity	Description
SKC-203A	Black Half Area 96 well Plate	One Plate	-
SKC-203B	Glutathione S-Transferase Standard	50 $\mu$ L	Equine Glutathione S-Transferase at 10 U/mL in a special stabilizing solution.
SKC-203C	GST Detection Reagent	1 vial	GST detection substrate stored in a desiccator.
SKC-203D	Dry DMSO	2 mL	Dry Dimethyl sulfoxide solvent over molecular sieves. May be stored at room temperature.
SKC-203E	Assay Buffer	45 mL	Phosphate buffer containing proteins and stabilizers.
SKC-203F	Glutathione (GSH)	300 $\mu$ L	Glutathione supplied as a 20 mM stable solution.

If any of the items listed above are damaged or missing, please contact our Customer Service department at (250) 294-9065. We cannot accept any returns without prior authorization.



**WARNING: Not for human or animal disease diagnosis or therapeutic drug use.**

## Precautions

As with all such products, this kit should only be used by qualified personnel who have had laboratory safety instruction. The complete booklet should be read and understood before attempting to use the product.

Dimethyl sulfoxide is a powerful aprotic organic solvent that has been shown to enhance the rate of skin absorption of skin-permeable substances. Wear protective gloves when using the solvent especially when it contains dissolved chemicals.

## Storage

All components of this kit should be stored at 4°C until the expiration date of the kit.

DMSO, when stored at 4°C, will freeze. Can be stored tightly capped at room temperature.

## Materials Needed But Not Supplied

- Repeater pipet with disposable tips capable of dispensing 25  $\mu$ L.
- Fluorescence 96 well plate reader capable of reading fluorescent emission at 460 nm, with excitation at 390 nm. Set plate parameters for a 96-well Corning Costar 3686 plate. Plate Dimensions (in mm): Well Depth: 10.54; Plate Length: 127.8; Plate Width: 85.5; A1 Row Offset: 11.2; A1 Column Offset: 14.3.
- Software for converting raw relative fluorescent unit (FLU) readings from the plate reader and carrying out four parameter logistic curve (4PLC) fitting. Contact your plate reader manufacturer for details.

## Activity Standardization

The Glutathione S-Transferase standard used in this kit has been calibrated using an enzymatic method adapted from reference 1.

**Please read this booklet completely prior to using the product.**

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

## Background

The Glutathione S-Transferase (GST) family of isozymes function to detoxify and neutralize a wide variety of electrophilic molecules by mediating their conjugation with reduced glutathione<sup>1</sup>. Human GSTs are encoded by 5 gene families, expressing in almost all tissues as four cytosolic and one microsomal forms. Dividing the family by isoelectric points, the basic alpha (pI 8–11), the neutral mu (pI 5–7) and acidic pi (pH<5) classes are populated by additional subclasses, each isozyme displaying differential specificity for given electrophilic molecules<sup>2</sup>.

Given its pivotal role in ameliorating oxidative stress/damage, GST activity has been repeatedly investigated as a biomarker for arthritis, asthma, COPD, and multiple forms of cancer, as well as an environmental marker<sup>3-7</sup>. Examination of GST isoforms and activity in human cancers, tumors and tumor cell lines has revealed the predominance of the acidic pi class. Furthermore, this activity is thought to substantially contribute to the innate or acquired resistance of specific neoplasms to anticancer therapy<sup>8,9</sup>.

## Assay Overview

The Glutathione S-Transferase *StressXpress*<sup>®</sup> Fluorescent Activity kit is designed to quantitatively measure the activity of GST present in a variety of samples. Please read the complete kit booklet before performing this assay. A GST standard is provided to generate a standard curve for the assay and all samples should be read off the standard curve. The kit utilizes a non-fluorescent molecule that is a substrate for the GST enzyme which covalently attaches to glutathione (GSH) to yield a highly fluorescent product. Mixing the sample or standard with the supplied Detection Reagent and GSH and incubating at room temperature for 30 minutes yields a fluorescent product which is read at 460 nm in a fluorescent plate reader with excitation at 390 nm. The activity of the GST in the sample is calculated, after making a suitable correction for any dilution of the sample, using software available with most fluorescence plate readers.

We have provided a 96 well plate for measurement but this assay is adaptable for higher density plate formats. The end user should ensure that their HTS black plate is suitable for use with these reagents prior to running samples.

## PRE-ASSAY PREPARATION

### Sample Types

**Sample Types Validated:**  
Serum, Plasma and Cell Lysates

This assay has been validated for human urine, serum, EDTA, heparin plasma, toadfish liver (*Opsanus tau*) and oyster hemolymph samples. Most cell lysates should also be compatible. Samples that are not clear or that contain visible particulate should be centrifuged prior to using.

GST activity varies across tissues and species, however we expect this kit to measure GST activity from sources other than human. The end user should evaluate recoveries of GST activity in samples from other species being tested.

### Sample Preparation

Any samples requiring larger dilutions or with GST activities outside the standard curve range should be diluted further with Assay Buffer to obtain readings within the standard curve.

### **Urine Samples**

Samples that are not clear or that contain visible particulate should be centrifuged prior to using. Urine samples should be diluted  $\geq 1:2$  in Assay Buffer by adding one part of urine to one part of Assay Buffer. Sample values should be normalized for urinary volume to urinary creatinine levels. Our Creatinine Urinary Detection Kit, SKT-200 allow simple, safe and accurate determination of urinary creatinine levels.



## Serum and Plasma Samples

Samples that are not clear or that contain visible particulate should be centrifuged prior to using. Fresh serum or EDTA and heparin plasma are separated by centrifugation at 600 x g for 10 minutes. Transfer the serum or plasma from the red blood cells into fresh tubes. The serum or plasma may be stored at -80°C or analyzed immediately. Serum or plasma should be diluted with assay Buffer at a dilution of  $\geq 1:2$ .

## Cell Lysates

Washed cell pellets are resuspended at  $10\text{-}40 \times 10^6$  cells/mL in Assay Buffer (we used Jurkats at  $10 \times 10^6$  cells/mL) and are lysed by vigorous vortexing, freeze/thaw cycling or other suitable disruption method. Resulting centrifuged lysate supernatants are measured at appropriate dilutions. The protocol might require adjustment for other cell types. If protein determinations are to be made on the samples, we would recommend using higher number of cells and lysing in your normal PBS-based lysis buffer and determining protein concentration, prior to additional dilutions in Assay Buffer to measure GST activity.

Use all samples within 2 hours of dilution.

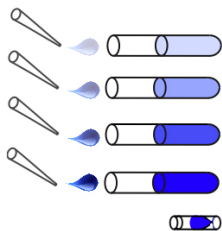
## Reagent Preparation

Allow the kit reagents to come to room temperature for 30 minutes. We recommend that all standards and samples be run in duplicate to allow the end user to accurately determine GST activities. Ensure that all samples have reached room temperature and have been diluted as appropriate prior to running them in the kit.

### **Standard Preparation**

GST Standards are prepared by labeling seven test tubes as #1 through #7. Briefly spin vial of standard in a microcentrifuge to ensure contents are at bottom of vial. Pipet 380  $\mu\text{L}$  of Assay Buffer into tube #1 and 200  $\mu\text{L}$  into tubes #2 to #7. Carefully add 20  $\mu\text{L}$  of the Glutathione S-Transferase Standard to tube #1 and vortex completely. Take 200  $\mu\text{L}$  of the GST solution in tube #1 and add it to tube #2 and vortex completely. Repeat these serial dilutions for tubes #3 through #7. The concentration of GST in tubes 1 through 7 will be 500, 250, 125, 62.5, 31.25, 15.61 and 7.81 mU/mL.

Use all Standards within 1 hour of preparation.



	Standard 1	Standard 2	Standard 3	Standard 4	Standard 5	Standard 6	Standard 7
Buffer Volume (μL)	380	200	200	200	200	200	200
Addition	Stock	Standard 1	Standard 2	Standard 3	Standard 4	Standard 5	Standard 6
Volume of Addition (μL)	20	200	200	200	200	200	200
Final Concentration (mU/mL)	500	250	125	62.5	31.25	15.625	7.81

## Detection Reagent

Remove the vial of Detection Reagent from the desiccator and add 300  $\mu\text{L}$  of the dry DMSO to the vial. Vortex thoroughly. Store any unused reconstituted Detection Reagent at 4°C in the desiccator for no longer than 2 weeks.

Dilute one part of reconstituted Detection Reagent 1:10 into nine parts of Assay Buffer. 150  $\mu\text{L}$  of Detection Reagent should be diluted with 1.35 mL of Assay Buffer to use half the plate. Discard any excess diluted Detection Reagent.

## Glutathione

Dilute one part Glutathione stock provided 1:10 into nine parts of Assay Buffer. 150  $\mu\text{L}$  of Glutathione stock should be diluted with 1.35 mL of Assay Buffer to have enough GSH to be able to read half the plate. Discard any excess diluted glutathione.

### Assay Protocol

1. Use the plate layout sheet on page 22 of the booklet to aid in proper sample and standard identification. Set plate parameters for a 96-well Corning Costar 3686 plate.
2. Pipet 50  $\mu\text{L}$  of treated samples or standards into duplicate wells in the plate.
3. Pipet 50  $\mu\text{L}$  of Assay Buffer into duplicate wells as the Zero standard.
4. Add 25  $\mu\text{L}$  of the Detection Reagent to each well using a repeater pipet.
5. Add 25  $\mu\text{L}$  of GSH to each well using a repeater pipet.
6. Gently tap the sides of the plate to ensure adequate mixing of the reagents.
7. Incubate at room temperature for 30 minutes.
8. Read the fluorescent signal from each well in a plate reader capable of reading the fluorescent emission at 460 nm with excitation at 370–410 nm. Please contact your plate reader manufacturer for suitable filter sets.
9. Use the plate reader's built-in 4PLC software capabilities to calculate GST activities for each sample.

## Calculation of Results

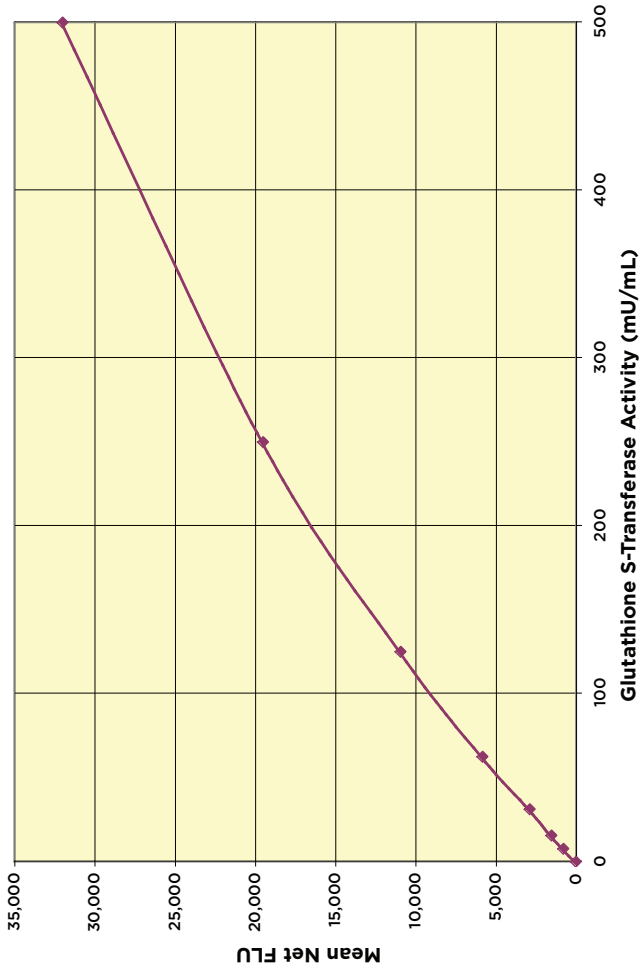
Average the duplicate FLU readings for each standard and sample. Create a standard curve by reducing the data using the 4PLC fitting routine on the plate reader, after subtracting the mean FLUs for the zero standard. The concentrations obtained should be multiplied by the dilution factor to obtain neat sample values.

## Typical Data

Sample	Mean FLU	Net FLU	GST Activity mU/mL
Zero	3,669	0	0
Standard 1	35,629	31,988	500
Standard 2	23,147	19,506	250
Standard 3	14,566	10,925	125
Standard 4	9,474	5,833	62.5
Standard 5	6,517	2,876	31.25
Standard 6	5,617	1,526	15.61
Standard 7	4,425	784	7.81
Sample 1	30,109	26,468	376.6
Sample 2	7,094	3,453	36.52

Always run your own standard curve for calculation of results.  
Do not use this data.

## Typical Standard Curve



## Validation Data

### **Sensitivity and Limit of Detection**

Sensitivity was calculated by comparing the FLUs for twenty wells run for each of the zero and standard #7. The detection limit was determined at two (2) standard deviations from the zero along the standard curve.

Sensitivity was determined as 2.70 mU/mL.

The Limit of Detection was determined in a similar manner by comparing the FLUs for twenty wells run for each of the zero and a low concentration serum sample.

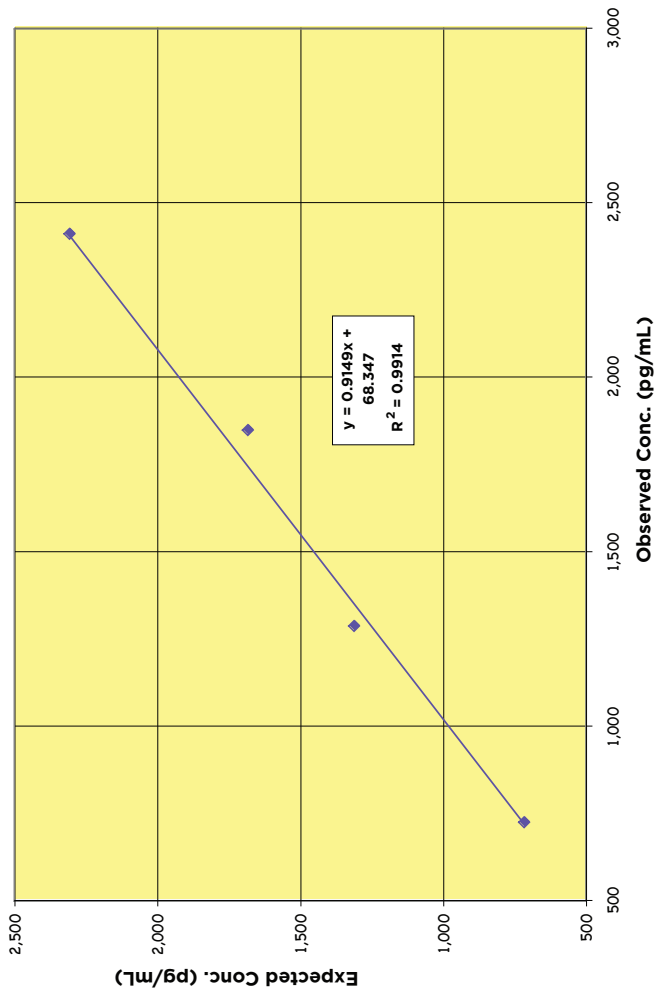
The Limit of Detection was determined as 1.90 mU/mL.

### **Linearity**

Linearity was determined by taking Jurkat cell lysates at  $10 \times 10^6$  and  $0.8 \times 10^6$  cells/mL and mixing in the ratios given below. The measured activities were compared to the expected values based on the ratios used.

Low Cell #	High Cell #	Expected Activity (mU/mL)	Observed Activity (mU/mL)	% Recovery
100%	0%	40.4	--	--
80%	20%	123.3	122.4	99.3
60%	40%	206.2	217.2	105.3
40%	60%	289.1	297.1	102.7
20%	80%	372.1	395.1	106.2
0%	100%	455.0	--	--
			Mean Recovery	103.4%





### Intra Assay Precision

Four serum samples were diluted 1:2 with Assay Buffer and run in replicates of 16 in an assay. The mean and precision of the calculated GST activities were:

Sample	GST Activity (mU/mL)	%CV
1	315.9	4.6
2	221.2	5.6
3	88.2	4.2
4	22.7	6.6

### Inter Assay Precision

Four serum samples were diluted 1:2 with Assay Buffer and run in duplicates in twenty assays over multiple days by four operators. The mean and precision of the calculated GST activities were:

Sample	GST Activity (mU/mL)	%CV
1	291.7	12.6
2	218.5	11.0
3	89.6	10.4
4	23.0	15.9

## Sample Values

Twenty random human serum, heparin and EDTA plasma samples were tested in the assay. Values ranged from 26.8 to 59.8 mU/mL with an average of 39.1 mU/mL. Five random human urine samples were tested in the assay and values were normalized to urinary Creatinine levels utilizing the StressMarq Biosciences Creatinine Urinary Detection kit, SKT-200. Levels ranged from almost 24 to over 102 mU/mL. When normalized for urine volume using creatinine values, the levels ranged from 2.83 to 20.2  $\mu$ U/mg creatinine.

## Interferences

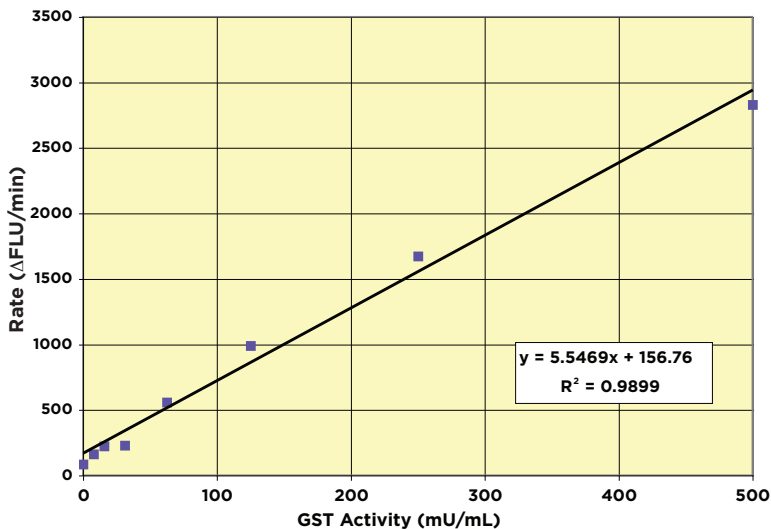
A variety of solvents and detergents were tested as possible interfering substances in the assay. Approximately 10% change was seen in the GST activity in the presence of 1% methanol or DMSO in the sample. Three detergents were also tested, Triton X-100, Tween 20 and SDS. At 0.01% concentration in the sample both SDS and Tween showed no change in activity, whereas Triton showed >47% decrease at 0.01%.

Bilirubin levels of 2.5  $\mu$ g/mL in the sample showed < 5% decrease in GST activity.

## End Point versus Kinetic Activity

The assay can also be run as a kinetic assay. A human serum sample was read in both an end point and in a kinetic assay. In the end point measurement it had a reading of 12.12 mU/mL and in the kinetic assay a reading of 11.92 mU/mL. A typical standard curve for the kinetic assay is shown below.

### Kinetic Assay



### References

1. Habig, W, et al. "Glutathion S-Transferases: The First Enzymatic Step in Mercapturic Acid Formation" *J. Biol.Chem.* 1974 249(22):7130-7139.
2. Cook, JA, et al., "Differential Specificity of Monochlorobimane for Isozymes of Human and Rodent Glutathione S-Transferases" *Cancer Res.* 1991 51:1606-1612.
3. Dalle-Donne, I, et al. "Biomarkers of Oxidative Damage in Human Disease" *Clinical Chemistry*, 2006 52(4):601-623.
4. Surapneni, KM & VSC Gopan, "Lipid Peroxidation and Antioxidant Status in Patients with Rheumatoid Arthritis" *Ind.J.Clin.Biochem.* 2008 23(1):41.44.
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8. Shea, TC, et al., "Identification of an Anionic Form of Glutathione Transferase Present in Many Human Tumors and Human Tumor Cell Lines" *Cancer Res.* 1988 48:527-533.
9. Shea, TC, et al., "Glutathione Transferase Activity and Isozyme Composition in Primary Human Breast Cancers" *Cancer Res.* 1990 50:6848-6853.

## Warranty and Limitation of Remedy

StressMarq Biosciences Inc. makes **no warranty or guarantee** of any kind, whether written or oral, expressed or implied, including without limitation, any warranty of fitness for a particular purpose, suitability and merchantability, which extends beyond the description of the chemicals hereof. StressMarq **warrants only** to the original customer that the material will meet our specifications at the time of delivery. StressMarq will carry out its delivery obligations with due care and skill. Thus, in no event will StressMarq have **any obligation or liability**, whether in tort (including negligence) or in contract, for any direct, indirect, incidental or consequential damages, even if StressMarq is informed about their possible existence. This limitation of liability does not apply in the case of intentional acts or negligence of StressMarq, its directors or its employees.

Buyer's **exclusive remedy** and StressMarq's sole liability hereunder shall be limited to a refund of the purchase price, or at StressMarq's option, the replacement, at no cost to Buyer, of all material that does not meet our specifications.

Said refund or replacement is conditioned on Buyer giving written notice to StressMarq within thirty (30) days after arrival of the material at its destination. Failure of Buyer to give said notice within thirty (30) days shall constitute a waiver by Buyer of all claims hereunder with respect to said material.

**For further details, please refer to our Warranty and Refund Policy located on our website and in our catalog.**

## Contact Information

<b>Phone:</b>	250-294-9065
<b>Fax:</b>	250-294-9025
<b>E-Mail:</b>	techsupport@stressmarq.com
<b>Hours:</b>	M-F 9:00 AM to 5:00 PM PST

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

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